

## ACCUMULATION OF HEAVY METALS IN WASTEWATER, SOIL AND UPTAKE BY PLANT SPECIES WITH PHYTOREMEDIATION POTENT

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### Abstract:-

The present study was to evaluate the concentrations of heavy metals (Cd, Cr and Cu) in soils, waste water and different plant organs; define which species and which plant organs exhibit the greatest accumulation and evaluate whether these species could be usefully employed in phytoremediation programs. Al- Marj plain is located in north eastern parts of Libya at the western edge of Al-Akhdar Mountain. Seven locations, distributed in polluted area. Data collected from heavy metal analysis demonstrated that the levels of the recorded metals in waste water were above the standard levels of USEPA, FAO and standard levels of Syrian Standard adopted by National Environmental Quality Standards for municipal and liquid industrial effluents in Libya. Heavy metals concentration in polluted soil samples were decreased in Sequence of: Cu>Cr>Cd. The thirteen species were subjected to suitable analysis to determine their capabilities for heavy metals accumulation as well as the metal allocation was also investigated. The results indicated that Cd could be accumulated in all plant organs (roots, stems and leaves). Generally, roots of some selected plants attained higher Cr and Cu, concentrations than other organs. The bioaccumulation factors of the investigated metals (BAF) increased in the sequence: Cu > Cr>Cd. According to the results *Conium maculatum*, *Juncus rigidus*, *Urtica dioica* and *Phragmites australis* could be used successfully in bio monitoring and phytoremediation programs for heavy metal contaminated soils.

**Keywords:-** Heavy metal, accumulation, Phytoremediation, Polluted soils bioaccumulation factor.

## INTRODUCTION

In developing countries, fast-growing urban populations are demanding more fresh water and food, while generating greater volumes of wastewater. Due to the lack of comprehensive wastewater management, a major portion of the wastewater pollutes natural water bodies [1]. These polluted sources are used in and around the cities for agriculture and other purposes. Soil pollution, is a very important environmental problem and it has been attracting considerable attention in recent years. Deteriorating soil quality and decrease in vegetation abundance are grave consequences of open waste dumping which have resulted in growing public concern [2]. In modern economies various types of activity including agriculture, industry and municipal produce a larger amount of wastes and new type of pollutants. Soil, air and water have traditionally been used as sites for disposal of all these wastes. Heavy metal contamination of soil is a major environmental problem that can reduce both the productivity of plant and safety of plant products as food and feed [3]. With the rise of global heavy metal contamination, the plants that can accumulate heavy metals might provide efficient and ecologically sound approaches to sequestration and removal of these pollutants [4]. Plants of a high metal bioaccumulation capacity and a good tolerance to high metal concentration over long periods of time could be efficient phytoremediator.

Since 30 years ago many places in Libya are subjected to untreated waste water discharge. Al-Marj plain is an example of such places. It has been impacted for many years by untreated industrial, agricultural and domestic waste waters through different discharges and drains [5]. This Study aims to: 1- Evaluate the concentrations of heavy metals (Cd, Cr and Cu) in soils, waste water and different plant organs. 2- Define which species and which plant organs exhibit the greatest accumulation and evaluate whether these species could be usefully employed in phytoremediation programs.

## Materials and methods

### Site characterization

The plant, waste water and soil samples used in this study were collected from a known waste water contaminated site located in Al-Marj plain, located on the southern coast of the Mediterranean Sea in the province of Cyrenaica [6], Cyrenaica houses Al-Jabal AlKhdar (the Green Mountain). The study area extends from Farzougha west to Al-Beyda city and bordered northeasterly by Batta and southeasterly by Gardis. It is penetrated by some agro- and populated areas [7]. Astronomical site extends between latitudes 33° and 31°N and longitudes 20.30°- 21.30°E. It depends on ground water of 200-350m deep as the chief source of the potable water [5]. In Al-Jabal Al-Akhdar area precipitation is the main factor that determines vegetation cover and land use. Traditionally, extensive dry farming cultivation of wheat and barley and stock-herding with goats and sheep were the main activities in Al-Jabal Al-Akhdar region [8]. Our observation showed that the vegetation was few and non-compact. Plant species collected were the most common/dominant species at the polluted area. A total of thirteen plants, waste water and soils (at 0-20 cm depth from roots of each plant were taken from each site from where plant samples were rooted) were collected in March 2013 in triplicate from studied locations for heavy metal determination and their scientific names and characteristics were determined.

### Sampling

Soils as well as thirteen abundant and dominating native plants (*Eucalyptus leucoxylon*, *Nicotianaglauca*, *Amaranthusviridis*, *Conium maculatum*, *Pinushalepensis*, *Phragmitesaustralis*, *Ceratoniasiliqua*, *Juncusrigidus*, *Cupressussempervirens* var. *horizontalis*, *Ficuscarica*, *Ricinuscommunis*, *Urticadioica* and *Foeniculumvulgare*) were collected from seven polluted locations in the study area for each soil sample heavy metals were measured. Plants sampling at least three whole plants of each species of current year were collected and then washed using tap water then distilled water to remove all the debris and other foreign particles, and then separated into stems, leaves and roots to identify the different accumulation capability and selectivity of each organ. The native plant species were identified according to [9]. For waste water samples were collected in polyethylene bottles About 1 liter waste water sample collected from each location in the study area. These samples were acidified immediately in the field with Nitric acid (1ml HNO<sub>3</sub>/L) for heavy metals determinations.

### Analytical techniques

Soil samples (a composite mixture) were air dried, then dried in a furnace at 120°C for 48 h. After drying, samples were cleaned off from any stones and plant residues, grounded in a stainless steel soil grinder then, passed through 2 mm sieve for heavy metal analysis, one gram of homogenized samples was digested using HNO<sub>3</sub>-HF-H<sub>3</sub>BO<sub>4</sub> acids according to [10]. Plant materials were oven-dried to a constant mass into oven at 80°C and grounded to a fine powder. In this way homogeneous samples were obtained for each plant organ. Approximately 0.2 gram of leaves, roots and stems powder were weighed and digested according to method described by [11]. Waste water samples were filtrated to remove suspended solids by glass fiber filters (Millipore type HA 0.45 μm pore size, from Millipore Corporation, Bedford, Massachusetts 01730, USA) using a vacuum pump according to standard procedures [12]. About 1 liter water sample collected from each location in the study area. These samples were acidified immediately in the field with Nitric acid (1ml HNO<sub>3</sub>/L) for heavy metals determinations. Soil, waste water and plant samples were analyzed for heavy metals by Perkin-Elmer 2380 atomic absorption spectrophotometer which calibrated with standard solutions containing known concentrations of each element. Standard solutions were prepared by diluting available high purity stock solutions (BDH). Average values of three replicates were taken for each determination. The precision of analytical procedures was expressed as relative standard deviation which ranged from 510% / was calculated from the standard deviation divided by the mean. Bioaccumulation factors (BAF) was calculated using the formula outlined by [13].

$$\text{BAF} = \frac{\text{element concentration in plant root}}{\text{element concentration in waste water}}$$

## RESULTES AND DISCUSSION

### Waste water, soil and plant analysis

Domestic waste water contain fairly high concentration of metals such as copper, iron, lead and zinc, which are derived from household products such as cleaning materials, toothpaste, cosmetics and human feces[14]. Also, there are additional quantities introduced (illegally into water canals) from industrial effluents. Most of the heavy metals are extremely toxic because of their solubility in water. In the present study, heavy metal concentrations in waste water samples were decreased in sequence of Cr> Cu> Cd. The same pattern was recorded for the studied metals in the polluted soil samples collected from the same area. Excessive accumulation of heavy metals in agricultural soils through waste water effluent, may not only result in soil contamination, but also affect food quality and safety [15]. Average Cd,Cr and Cu concentration of waste water in the seven polluted locations are given in( Table 1).

Cadmium (Cd) is one of the most toxic heavy metal and is considered non-essential for living organisms [16].Plant treated with higher concentrations of Cd usually become stunted in growth [17]. Data collected from the present study showed that waste water Cd level exceeded the upper limit of the [18 and 19]( 0.02,0.01and0.01  $\mu\text{gml}^{-1}$ respectively), The maximum Cd waste water concentration was attained at location 4 (0.87  $\mu\text{gml}^{-1}$ ).

Chromium (Cr) is one of the toxic metals widely distributed in nature. It has two forms found in the environment, trivalent and hexavalent. The latter form is considered to be the greatest threat because of its strong oxidizing ability as well as high solubility and availability to penetrate cell membranes [20]. Results obtained from this study, showed that Cr concentrations in waste water were higher than 0.73 $\mu\text{gml}^{-1}$  in all the investigated polluted locations .Thus exceeds the upper limit of [18and 19] (0.074, 0.1and 0.1 $\text{gml}^{-1}$  respectively). Copper (Cu) is a micronutrient for plants at very low concentration. However, excessive c

On centration of this metal are considered to be highly toxic. Mining, smelting and land application of fertilizers and sewage sludge together with the use of fungicides containing Cu and other human activities has led to wide spread soil contamination with Cu[21].Results obtained from this study, showed that Cu concentrations in waste water were higher than 0.40 $\mu\text{gml}^{-1}$  in all the investigated locations. Thus exceeds the upper limit of [18 and 19] (0.013 0.2 and 0.2  $\mu\text{gml}^{-1}$  respectively).Cu speciation and toxicity in water may be influenced by a variety of physicochemical variables, particularly water hardness, alkalinity, pH and natural organic matter [22].

According to the results of this study, the native plants and soil can well present further information about the metal content of their environment. Average soil Cd, Cr and Cu concentration of the seven polluted locations are given in (Table 2). Average content of Cd in soils lies between 0. 06 - 1.1  $\mu\text{gg}^{-1}\text{d.w}$ . [23]. The average soil Cd concentration recorded in the current study for the polluted locations was relatively high, 4.37 $\mu\text{gg}^{-1}\text{d.w}$ . Cd is considered as a heavy metal of most concern because it shows the greatest mobility in the soil environment [24] . The main anthropogenic sources of Cd are the amendment of agricultural soils with Cd contaminated bio-solids, phosphate fertilizers and industrial biproducts [25].In the current investigation, values of extractable Cd of all polluted soils samples were higher than limit values in soils (1-3  $\mu\text{gg}^{-1}\text{d.w}$  ) according to the Council of the European Communities[26]. except for locations 2 and 7 (2.95 and 2.66  $\mu\text{gg}^{-1}\text{d.w}$  respectively).Sewage sludge, smelting operation and phosphate fertilizers are important sources of higher Cd concentration in the topsoil[23].Generally, the highest Cr concentration of soil samples collected from the polluted locations(46.70 $\mu\text{gg}^{-1}\text{d.w}$ ) was recorded at location 3. This location is directly affected by the discharge of industrial effluents of a nearby textile and garments factories as well as municipal sewage sledges. While Cu concentration in the soils of the studied location lies within the accepted range (50-140  $\mu\text{gg}^{-1}\text{d.w}$ ) of soil Cu content as reported by Council of the European Communities [26].However, the investigation native plants exhibited different element concentrations depending on plant organ and the sampling site.Results also indicated that Cd could be accumulated in all plant organs (root, stem and leaves). The distribution of Cd within plant organs is quite variable and clearly illustrates its rapid translocation from roots to shoots [23]. According to [27], Cd content of most plants did not exceed 1.9 $\mu\text{gg}^{-1}\text{d.w}$ . However, the Cd content recorded for all the studied species of the polluted locations exceeded this threshold and reached up to 22.78  $\mu\text{gg}^{-1}\text{d.w}$ . *Eucalyptus leucoxylo*n stem are given in Fig.1.

Results from the present study showed that the maximum Cr concentration in all the investigated plant species was always contained in the root system. The highest Cr concentration was attained by the roots of *Ceratonia siliqua* at location 7 (395.43  $\mu\text{gg}^{-1}\text{d.w}$ ). This is in agreement with[28 and 29] .This could be because Cr is immobilized in the vacuoles of the root cells and showed less translocation, thus rendering it less toxic. According to [30] the toxic levels of Cr in plants range from 1 to 10  $\mu\text{gg}^{-1}$  dry weight .Cr content of the weight .Cr content of the studied species could be arranged according to the pattern:

*Amaranthusviridis*>*Coniummaculatum*>*Phragmitesaustralis*>*Eucalyptusleucoxylo*n>*Pinushalepensis*>*Foeniculumvulgare*>*Nicotianaglauca*>*Ficuscarica*>*Cupressussempervirens*var.*horizontalis*>*Ricinuscommunis*>*Juncusrigidus*>*Urtica dioica*>*Ceratonia siliqua* were given in Fig.2

Generally, roots of most of the studied plants in the polluted locations attained higher Cu concentration than other organs with maximum value of 578.15  $\mu\text{gg}^{-1}\text{d.w}$  attained by *Phragmitesaustralis* root. On the other hand, leaves and stems of both *Cupressussempervirens* and *Pinushalepensis* were found to accumulate considerable amounts of Cu (> 350  $\mu\text{gg}^{-1}\text{d.w}$ )(Fig.3). This is agreement with [28].Cu concentrations in plant tissue above 10-30  $\mu\text{gg}^{-1}\text{d.w}$  are regarded as poisonous [30]. Within roots, Cu is associated mainly with cell walls and is largely immobile. However, higher concentration of Cu in shoots (leaves and stems) are always in phases of intensive growth and at the luxury Cu supply level[31].High concentrations of Cu in the roots of *Phragmitesaustralis* with relatively high pH values in soil (> 7.0) may be attributed to the presence of plaque ,a metalrich rhizo -concentrations composed of iron hydroxides and other metals that are mobilized and precipitated on the root surface[32].

### Bioaccumulation in plants

Accumulation of metal ions varied greatly among plant species and uptake of an element by a plant is primarily depended on the plant species, its inherent controls and the soil quality [33]. Large number of factors control metal accumulation and bioavailability associated with soil and climatic conditions, plant genotype and agronomic management, including: active/passive transfer process, sequestration and speciation, redox states, the type of plant root system and the response of plants to elements in relation to seasonal cycles [34]. The bioaccumulation factor (BAF) of the investigated metals, measured in comparison with the total content of metals in water, is a parameter used to describe the transfer of trace elements from polluted water to plant tissue. However, the bioaccumulation factors (BAF) increased in the sequence: Cu > Cr > Cd for most of the species in almost all location. Most of the plants under investigation had BAF > 1. Current results also revealed that *Conium maculatum* acquired the highest BAF value for Cd, Cu, (1,112.58-1,009.41) followed by *Juncus rigidus*. However, the latter species acquired the second highest BAF value for Cr and Cu (310.09872.37). *Phragmites australis* is characterized by having the highest BAF value for Cr. *Urtica dioica* acquired the highest BAF value for Cd (Table 3). Heavy metals tolerant species with high BAF can be used in phytoremediation programs of contaminated soils as these species retains metals in their roots and limit metal mobility from roots to shoots once absorbed by roots of plants [35]. According to the results collected from this investigation, the studied native plants and soil can well present further information about the metal content of their environment. Moreover, *Conium maculatum*, *Juncus rigidus*, *Phragmites australis* and *Urtica dioica* could be used successfully in bio-monitoring and phytoremediation programs for heavy metal contaminated soils.

**Table 1 .Average concentrations (mean± SD) of the investigated heavy metals in waste water samples ( $\mu\text{gml}^{-1}$ ) collected from the studied polluted locations. Highest and lowest values are underlined.**

Location	Cd	Cr	Cu
1	0.40 ± 0.02	0.97 ± 0.14	0.72 ± 0.02
2	0.69 ± 0.05	1.01 ± 0.06	0.53 ± 0.17
3	0.46 ± 0.05	0.81 ± 0.17	0.57 ± 0.05
4	<u>0.87 ± 0.04</u>	0.84 ± 0.06	<u>0.43 ± 0.11</u>
5	<u>0.26 ± 0.08</u>	<u>0.73 ± 0.08</u>	<u>0.82 ± 0.04</u>
6	0.79 ± 0.05	<u>1.44 ± 0.02</u>	0.64 ± 0.11
7	0.73 ± 0.10	0.78 ± 0.16	0.48 ± 0.10

**Table 2. Average concentration (± SD) of the investigated heavy metals in the soil samples ( $\mu\text{g}^{-1}\text{d.w}$ ) collected from the polluted studied locations. Highest and lowest values are underlined.**

Locations	Concentration ( $\mu\text{g}^{-1}\text{d.w}$ )		
	Cd	Cr	Cu
1	5.06 ± 0.60	32.30 ± 0.67	67.24 ± 0.25
2	2.95 ± 0.66	41.28 ± 1.80	<u>42.33 ± 0.66</u>
3	4.78 ± 0.55	<u>46.70 ± 1.40</u>	71.85 ± 0.49
4	<u>6.55 ± 0.44</u>	32.81 ± 1.26	77.11 ± 0.02
5	5.35 ± 0.52	42.16 ± 0.60	<u>88.27 ± 1.45</u>
6	3.25 ± 0.43	<u>31.90 ± 0.95</u>	60.92 ± 0.92
7	<u>2.66 ± 0.46</u>	38.14 ± 0.88	53.07 ± 0.17

**Table 3. Bio-accumulation factor (BAF) of Cd, Cr, Cu, in the selected species collected from the studied locations in the polluted site. Highest and lowest values are underlined.**

Locations	Species	Cd	Cr	Cu
1	<i>Amaranthusviridis</i>	29.40	293.67	329.21
	<i>Conium mculatum</i>	14.70	142.14	536.52
	<i>Foeniculumvulgare</i>	<u>4.76</u>	97.01	99.53
	<i>Ricinuscommunis</i>	5.21	76.83	122.33
2	<i>Conium mculatum</i>	19.50	138.62	598.45
	<i>Eucalyptus leucoxylon</i>	59.43	115.74	501.36
	<i>Foeniculumvulgare</i>	4.88	133.77	147.96
	<i>Ricinuscommunis</i>	7.62	83.64	121.10
3	<i>Amaranthusviridis</i>	29.83	230.49	702.20
	<i>Conium mculatum</i>	12.68	189.96	843.78
	<i>Nicotianaglauca</i>	24.66	73.10	58.44
	<i>Pinushalepensis</i>	10.12	108.51	620.64
4	<i>Conium mculatum</i>	11.71	306.73	590.17
	<i>Eucalyptus leucoxylon</i>	24.40	218.96	290.26
	<i>Nicotianaglauca</i>	22.96	158.71	<u>41.84</u>
	<i>Pinushalepensis</i>	14.48	180.35	454.13
5	<i>Conium mculatum</i>	<u>1,112.58</u>	259.27	345.96
	<i>Ficuscarica</i>	862.34	170.15	87.92
	<i>Juncusrigidus</i>	687.74	107.96	141.02
	<i>Urticadioica</i>	1,071.88	244.78	85.14
6	<i>Conium mculatum</i>	10.08	167.97	654.28
	<i>Cupressussempervirensvar. horizontalis</i>	14.29	<u>60.59</u>	757.06
	<i>Ceratoniasiliqua</i>	15.05	103.40	331.04
	<i>Phragmitesaustralis</i>	15.58	172.51	508.26
7	<i>Ceratoniasiliqua</i>	16.29	190.88	441.39
	<i>Conium mculatum</i>	15.47	111.86	<u>1,009.41</u>
	<i>Juncusrigidus</i>	10.91	310.09	872.37
	<i>Phragmitesaustralis</i>	16.86	<u>318.48</u>	677.67

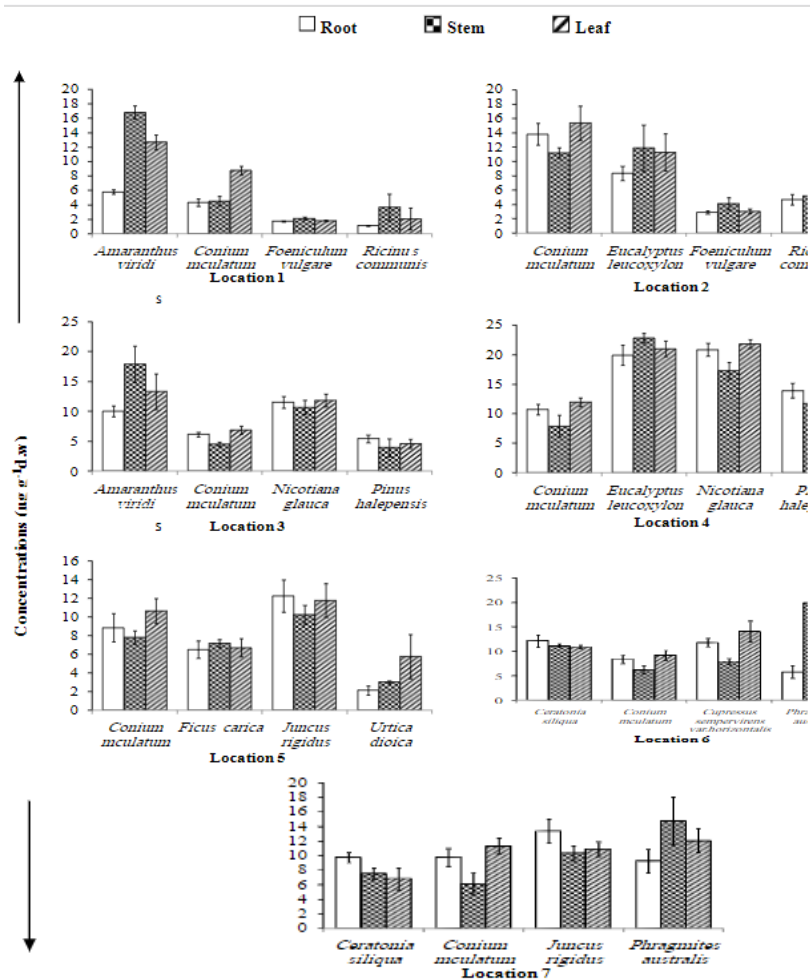
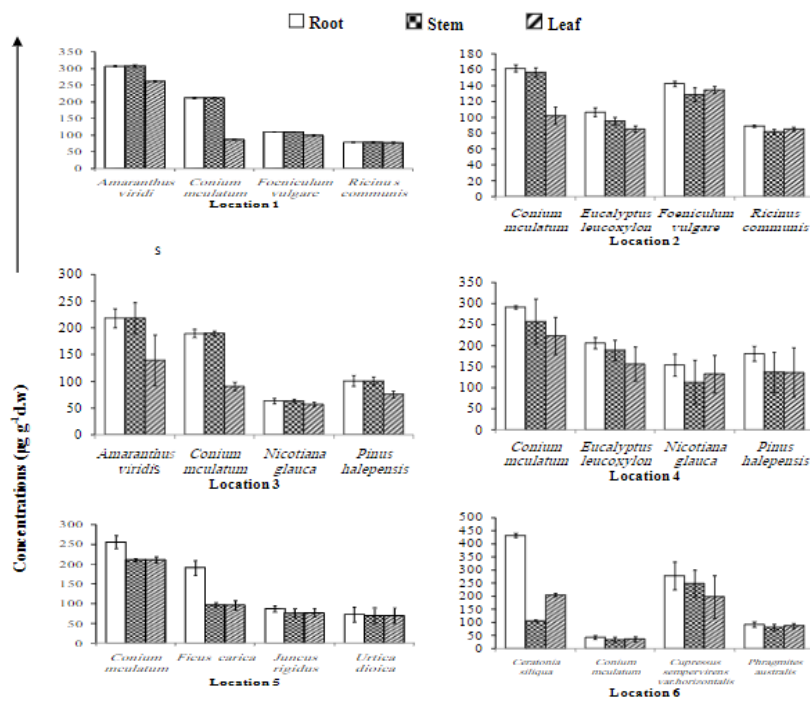


Fig.1. Average concentration of Cd in different organs of the selected plant species collected from the polluted studied locations together with standard deviation (SD) bars.



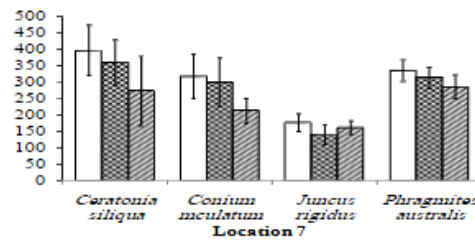


Fig. 2. Average concentration of Cr in different organs of the selected plant species collected from the polluted studied locations together with standard deviation (SD) bars.

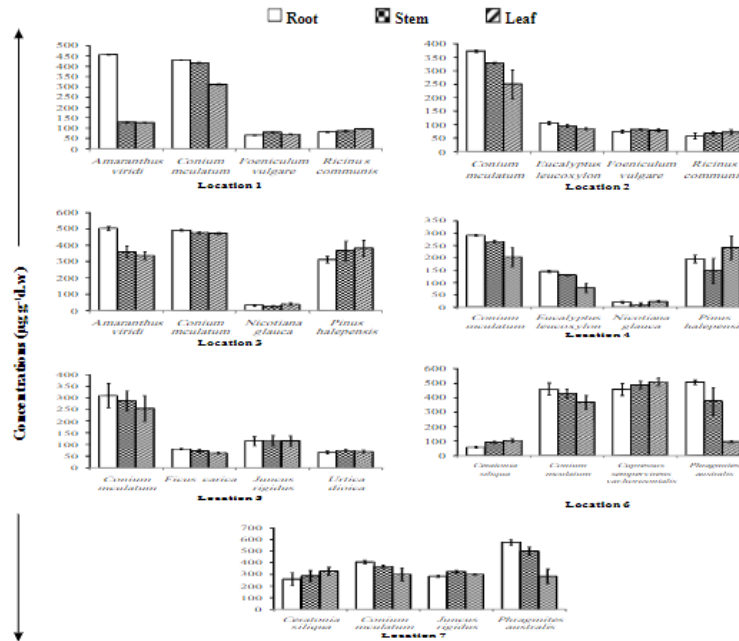


Fig.3. Average concentration of Cu in different organs of the selected plant species collected from the polluted studied locations together with standard deviation (SD) bars

**Recommendation:**

Serious actions to be taken to halt pollution by industrial and domestic waste water in AlMarj plain, and to the *Conium maculatum*, *Phragmites australis*, *Juncus rigidus* and *Urtica dioica* could be successfully used in biomonitoring and phytoremediation programs of polluted soils.

**References**

- [1]. Scott, C.A; Faruqi, N.I., and Raschid-Sally, L. (2004). Wastewater Use in Irrigated Agriculture: Confronting the Livelihood and Environmental Realities, CABI/IWMI/IDRC.
- [2]. Ali, S.M.; Pervaiz, A.; Afzal, B.; Hamid, N., and Yasmin, A. (2013). Open dumping of municipal solid waste and its hazardous impacts on soil and vegetation diversity at waste dumping sites of Islamabad city. Journal of King Saud University– Science. <http://dx.doi.org/10.1016/j.jksus.2013.08.003>.
- [3]. Tanhan, P.; Kruatrachue, M.; Pokethitiyook, P., and Chaiyarat, R. (2007). Uptake and accumulation of cadmium, lead and zinc by Siam weed [Chromolaena odorata (L.) King and Robinson]. Chemosphere, vol. 68, pp: 323-329.
- [4]. Rugh, C.L.; Wilde, H.D.; Stack, N.M.; Thompson, D.M.; Summers, A.O., and Meagher, R.B. (1996). Mercuric ion reduction and resistance in transgenic Arabidopsis thaliana plants expressing a modified bacterial merA gene. Proc Natl Acad Sci USA, vol. 93. No8, pp: 3182-8187.
- [5]. Alshamikh, A. (2009). Microbial pollution of the drink water in ElMarj city. Unpublished M.Sc. Thesis in Environmental science and Engineering at the Academy of Graduate Studies- Benghazi Branch, pp: 135.
- [6]. Newport, T.G., and Haddor, Y. (1963). Ground Water Exploration in Al-Marj Area, Creneica, Libya: Contribution to the Hydrology of Africa and the Mediterranean Region. United States Government Printing Office, Washington. pp 29.
- [7]. Alhassi, S.M. (2005). Effects of irrigation water quality on some physical and chemical properties of soils of four vegetable farms in Al-Marj basin. Unpublished M.Sc. Thesis in Environmental science and Engineering at the Academy of Graduate Studies- Benghazi Branch, pp: 157.
- [8]. Joffe, E.G.H. (1989). Libya-regional history, regional national borders. In Allan, J.A., McLachlan, K.S. and Buru, M. (eds) Libya: state and region, a study of regional evolution. London. Pp: 1-18.

- [9]. Jafri, S.M.H., and El-Gadi, A. (1979). *Flora of Libya*. vol.1-150 Al-faateh University, Faculty of Science Department of Botany, Tripoli. 15th ed. Constable and company. London, pp 498.
- [10]. Wade, T.L.; Brooks, J.M.; Kennicutt, M.C.; McDonald, T.J.; Sericano, J.L., and Jackson, T.L. (1993). GERG trace metals and organic contaminants analytical techniques. In: Lauenstein, G.G. and Cantillo, A.Y. (eds) *Sampling and analytical methods of the national status and trend program. National Benthic surveillance and mussel watch projects 1984-1992*, pp. 121-139. NOAA technical memorandum NOS ORCA 71. Silver Spring, MD.
- [11]. Allen, S.E. (1989). *Chemical analysis of Ecological Materials*. 2nd ed., Blackwell scientific publications, Oxford, pp: 430.
- [12]. A.P.H.A. (1985). *Standard methods for examination of water and waste*, 16th ed. Washington D.C.
- [13]. Sadler, R., and Rynja, G. (1992). *Preservation, Storage, Transport, Analysis and Reporting of water samples*, Queensland Government Chemical Laboratory Report Series No.12, Queensland Government publishers, Brisbane.
- [14]. Stephenson, T. (1987). Sources of heavy metals in wastewater. In: *Heavy metals in wastewater and Sludge treatment. Sources, Analysis and Legislation*. JN Lester CRC Press, Cleveland, OH. vol. 1. pp:31-64.
- [15]. Muchuweti, M.; Birkett, J.W.; Chinyanga, E.; Zvauya, R.; Scrimshaw, M.D., and Lester, J.N. (2006). Heavy metal content of vegetables irrigated with mixture of wastewater and sewage sludge in Zimbabwe: implications for human health *Agriculture, Ecosystem and Environment*, vol. 112. pp: 41–48 .
- [16]. Woodbury, P.B. (1998). *Municipal Solid Waste Composting: Potential Effects of heavy metals in Municipal Solid Waste Composts on Plants and the Environment*. Boyce Thompson Institute for Plant Research, Cornell University, Ithaca, NY.
- [17]. Wang, X.F., and Zhou, Q.X. (2005). Ecotoxicological effects of cadmium on three ornamental plants. *Chemosphere*, vol. 60. No1. pp: 16-21.
- [18]. USEPA (U.S. Environmental protection Agency). (2006). *National Recommended of water Quality Criteria*. Office of Water, Washington, DC.
- [19]. FAO (Food and Agriculture Organization). (1985). *Water Quality for Agriculture. Irrigation and Drainage*, United Nations, Rome. Paper No. 29, Rev. 1.
- [20]. Lytle, C.M.; Lytle, F.W.; Yang, N.; Qian, J.H.; Hansen, D.; Zayed, A., and Terry, N. (1998). Reduction of Cr (VI) to Cr (III) by Wetland plants: potential for in-situ heavy metal detoxification. *Environ Sci Technol.*, vol. 32. pp:3087-93.
- [21]. Levi-Minzi, R., and Riffaldi, R. (1978). Ricerche preliminary sul contenuto in metallic pesantideifanghi di depurazione. *Agricol. Ital.*, vol. 107. pp: 169-178.
- [22]. Niyogi, S., and Wood, C.M. (2004). Biotic ligand model, a flexible tool for developing site-specific water quality guidelines for metals. *Environ. Sci. Technol.*, vol. 38. pp: 61776192.
- [23]. Kabata-Pendias, A., and Pendias, H. (2001). *Trace Elements in Soils and Plants*. CRC Press, Boca Raton FL, USA.
- [24]. Wilson, M.J., and Bell, N. (1996). Acid deposition and heavy metal mobilization. *Appl. Geochem*, vol. 11. pp:133-137
- [25]. McLaughlin, M.J.; Tiller, K.G.; Naidu, R., and Stevens, D.P. (1996). Review: The behavior and environmental impact of contaminants in fertilizers. *Aust. J. Soil Res.*, vol. 34. pp: 1-54.
- [26]. C.E.C. (Council of the European Communities). (1986). Directive of 12 June 1986 on the protection of the environment, and in particular of the soil, when sewage sludge is used in agriculture (86/278/CEE). *Official Journal of the European Communities*, vol.181. pp: 612.
- [27]. Outridge, P.M., and Noller, B.N. (1991). Accumulation of toxic trace elements by freshwater vascular plants. *Reviews of Environmental Contamination and Toxicology*, vol. 121. pp: 1-63.
- [28]. Badr, N., Fawzy, M., and Al-Qahtani, K.M. (2012). Phytoremediation: An Ecological Solution to Heavy- Metal-Polluted Soil and Evaluation of Plant Removal Ability. *World Applied Sciences Journal*, vol. 16. No 9. pp: 1292-1301.
- [29]. Fawzy, M.A., and Badr, N.E. (2006). "Assessment of heavy metals accumulation capability of four aquatic plants in lake Mariut, Egypt." *Proceedings of the 2nd International Conference on Environmental Science and Technology*, Houston, USA. vol. 2. pp: 249-257.
- [30]. Macnicol, R.D., and Becktt, P.H.T. (1985). Critical Tissue Concentrations of Potentially Toxic Elements. *Plant Soil*, vol. 85. pp: 107-114.
- [31]. Tiffin, L. O. (1977). The form and Distribution of Metals in Plants: An Overview. In *Proc. Hanford Life Sciences Symp. U. S. Department of Energy, Symposium Series*, Washington, D. C. pp:315.
- [32]. Sundby, B.; Vale, C.; Cacador, I., and F. Catarino. (1998). Metal – rich Concentrations on The Roots of Salt Marsh Plants: Mechanism and Rate of Formation. *Limn. Oceanogr.*, vol. 43. pp: 245- 252.
- [33]. Chunilall, V., Kindness A. and Jonnalagadda, S.B. (2005). Heavy metal uptake by two edible *Amaranthus* herbs grown on soils contamination with Lead, Mercury, Cadmium and Nickle. *J. Environ. Sci. Health*, vol. 40. pp: 375-384.
- [34]. Kabata-Pendias, A., and Pendias, H. (1984). *Trace Elements in soil and plants*. CRC Press, Boca Raton, FL.
- [35]. Cui, S., Zhou, Q., and Chao, L. (2007). Potential hyper- accumulation of Pb, Cu, and Cd in enduring plants distributed in an old smeltery, northeast China. *Environ. Geol.*, vol. 51. pp: 1043–1048.